

Fluorinated Amino Acids
Part 3:¹ Synthesis of β -Difluoromethyl-*m*-tyrosine

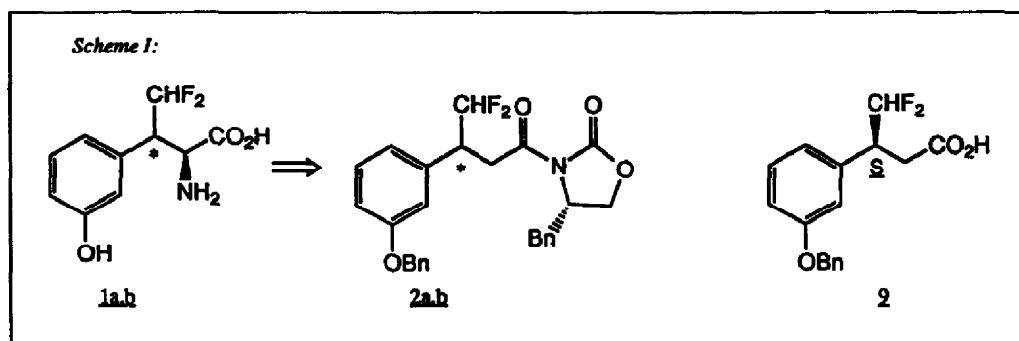
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Abstract: The difluoromethyl carboxylic acid S -(+)-**9** was synthesized to assign the absolute configuration of diastereomers **2a** and **2b**, and the completion of the synthesis of β -difluoromethyl-*m*-tyrosines **1a,b** is reported.

In the preceding letter, we detailed our approach to the synthesis of diastereomeric *N*-acyloxazolidinones **2a,b**, intermediates in the proposed synthesis of β -difluoromethyl-*m*-tyrosines **1a,b** (Scheme I). In this communication we disclose an independent asymmetric synthesis of carboxylic acid S -(+)-**9** which establishes the absolute configuration of the stereocenters in **2a,b**, and we describe the completion of the synthesis of **1a,b**.



A modification of our original approach¹ was used to prepare **9** (Scheme II). Commercially available aldehyde **3** was transformed through a geminal dibromoolefin to a terminal lithiated acetylide,² which was acylated with a Weinreb amide³ producing ketone **4**. Asymmetric reduction of **4** with *R*-Alpine-Borane⁴ afforded *R*-alkynyl alcohol **5** ($[\alpha]_D^{20} = -3.40^\circ$ ($c=1.0$, CHCl_3)) in 92% yield and 91% enantiomeric excess (ee) as determined by ¹⁹F NMR analysis of the Mosher ester.⁵ Reduction with lithium aluminum hydride⁶ (LAH) yielded an (*E*)-allylic alcohol (91% ee), $[\alpha]_D^{20} = -5.5^\circ$ ($c=0.8$, CHCl_3) which underwent an orthoester Claisen rearrangement with chirality transfer⁷ to afford ester **6** ($[\alpha]_D^{20} = -12.1^\circ$ ($c=1.03$, CHCl_3)). Ozonolysis of **6** then produced aldehyde **7**; treatment of **7** with diethylaminosulfur trifluoride (DAST)⁸ provided a sample of *gem*-difluoride S -(+)-**9** which was saponified to acid S -**9** ($[\alpha]_D^{20} = +23.2^\circ$ ($c=0.5$, CHCl_3)).

The final phase of the synthesis of **1a,b** involves the assignment of the absolute configuration of the stereocenters of **2a,b** by correlation with **9**, and the stereoselective introduction of the amino group. The sign of rotation of **S**-(+)-**9** is the same as that of the previously reported carboxylic acid ($[\alpha]_D^{20} = +39.2^\circ$ ($c=1.02$, CHCl_3)),¹ obtained by removal of the chiral auxiliary from diastereomer **2a**. By correlation therefore, the absolute configuration of the stereocenter in **2a** must be **S** and that of **2b** is **R**.

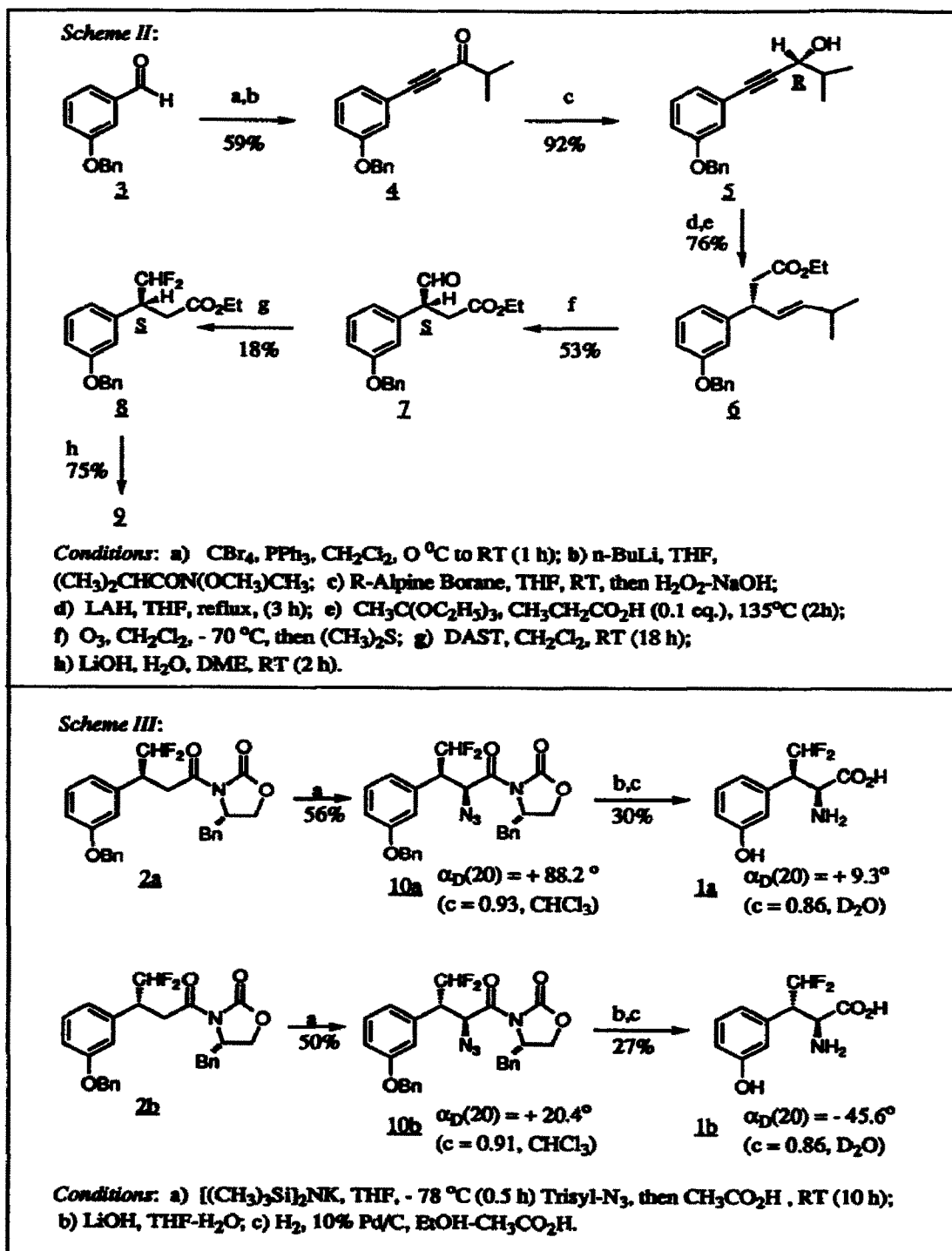
For the introduction of the amino group (Scheme III), the chiral imide enolates of **2a** and **2b** were quenched by electrophilic azide transfer with 2,4,6-triisopropylbenzene sulfonyl azide (Trisyl-N₃) ; good yields of α -azido carboximides **10a** and **10b** were obtained as single diastereomers after chromatography and ¹H NMR (300 MHz, CDCl₃) analysis. The diagnostic -CHF₂ signals at δ 6.26 (td, $J=2.9, 55.8$ Hz) for **10a** and δ 6.04 (td, $J=4.3, 56.2$ Hz) for **10b** were clearly the only ones observed, and diastereoselectivity was thereby judged to be complete. The relative stereochemistry of **10a** and **10b** was assigned based on *si*-face delivery of the azide group.¹⁰ Removal of the chiral auxiliary from **10a** and **10b** was effected using lithium hydroxide. Spectral analysis of the α -azido acids confirmed only single diastereomers. Catalytic hydrogenation (10% Pd/C) with concomitant hydrogenolysis of the benzyl ether completed the synthesis of **1a** and **1b**. Purification by HPLC (reverse phase)¹¹ afforded samples of diastereomerically pure **1a** and **1b**.¹²

In conclusion, our facile route to fluorinated amino acids **1a** and **1b** combines the synthesis and chromatographic separation of diastereomeric *N*-acyloxazolidinones **2a** and **2b** with the use of Evans' methodology for the electrophilic azidation of chiral imide enolates, and affords products of high diastereomeric purity for biological evaluation.

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References and Notes:

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2. Corey, E.J.; Fuchs, P.L. . *Tetrahedron Lett.*, **1972**, *13*, 3769.
3. For a review of applications in synthesis see: Sibi, M.P. *Org. Prep. Proc. Int.*, **1993**, *63*, 57.



4. For a discussion of the transition state leading to this enantioselectivity see: Midland, M. Organic Syntheses, 1984, **63**, 57.
5. Mosher, H.S.; Dale, J.A.; Dull, D.L. J. Org. Chem., 1969, **34**, 2543.
6. Corey, E.J.; Katzenellenbogen, J.A.; Posner, G.H. J. Am. Chem. Soc., 1967, **89**, 4245.
7. For a discussion of chirality transfer through sigmatropic rearrangements see: Hill, R.K. In Asymmetric Synthesis; Morrison, J.D.; Ed., Academic Press, Inc.: Orlando, FL 1984; Vol. III, p. 503.
8. Hudlicky, M. Org. React. (N.Y.), 1988, **35**, 513.
9. The sluggishness and poor yield for this transformation would suggest a possible erosion of chirality at this point, but for our purposes, provided an enriched sample of S-(+)-**8**. For a preparatively useful, multistep alternative see **5** $\xrightarrow{4 \text{ steps}}$ **7**, accompanying paper in this issue.
10. For a discussion of the kinetic diastereoselection of this step see: Evans, D.A.; Britton, T.C.; Ellman, J.A., Dorow, R.L., J. Am. Chem. Soc., 1990, **112**, 4011.
11. Purification was done using a Whatman Magnum 9 SCX (250 mm x 9.4 mm) column, with a mobile phase of 0.02M formic acid in water and a flow rate of 4 ml/min. RT (**1a**)=10 min; RT (**1b**)=16.5 min. Purified samples were reinjected on to a Partisil 10 SCX (250 mm x 4.6 mm.) column, same mobile phase, flow rate of 2 ml/min., and judged to be of >98% diastereomeric purity. We gratefully acknowledge the expertise of Mr. Nicholas W. Brake of our analytical chemistry department.
12. All structural assignments in the text are consistent with spectral and analytical data.

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